Read Alignment

Computational Problem

Given

- a reference genome G, and
- a set of reads $R = (r_1, r_2, r_3, ..., r_k) \in (\Sigma^n)^k$ where each read r is a subsequence of G with a small number changes

Output

• the semi-global alignment of r_i and G for all $r_i \in R$

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• the semi-global alignment of r_i and G for all $r_i \in R$ with < k changes

call these *k*-error mappings

Read Filtering

If a read is long enough, it should not align well to a random region of G

This assumes that the sequence was read correctly

Sequencing machines output a quality score for each position of a read

- this can be interpreted as a probability P(r[j]) that the character is correct
- in other words with probability P(r[j]), position r[j] is a random character

This means that a given sequence will match a random sequence with probability

$$\mathbb{P}(r) = \prod_{1 \le i \le n} \left(\mathbb{P}\left(r\left[j\right]\right) q_{r[j]} + \left(1 - \mathbb{P}\left(r\left[j\right]\right)\right) \right)$$

• where q_c is the probability of c in a random sequence

Read Filtering

We expect the number of random matches between r and a given string T such that |T| = m to be $\mathbb{E}(r, T) = (m - n + 1)\mathbb{P}(r)$

We can then threshold the reads that are too low quality by this expectation

Pigeonhole Principle

Assume for now we're not dealing with insertions or deletions

The **pigeonhole** principle in this case says that if the read is partitioned into k+1 pieces, one must appear in the genome exactly if the read has a k-error mapping.

All *k*-error mappings will have at least one exact match, not all exact matches lead to *k*-error mappings

Initial ideas to read mapping

Construct an index of G (say a succinct suffix array)

Search for each of the pieces of the read in the index

Verify each occurrence's alignment against that region of G

 $O(m \log \sigma + (\log^{(1+\varepsilon)}n + m^2/w) c)$ time

Aligning reads

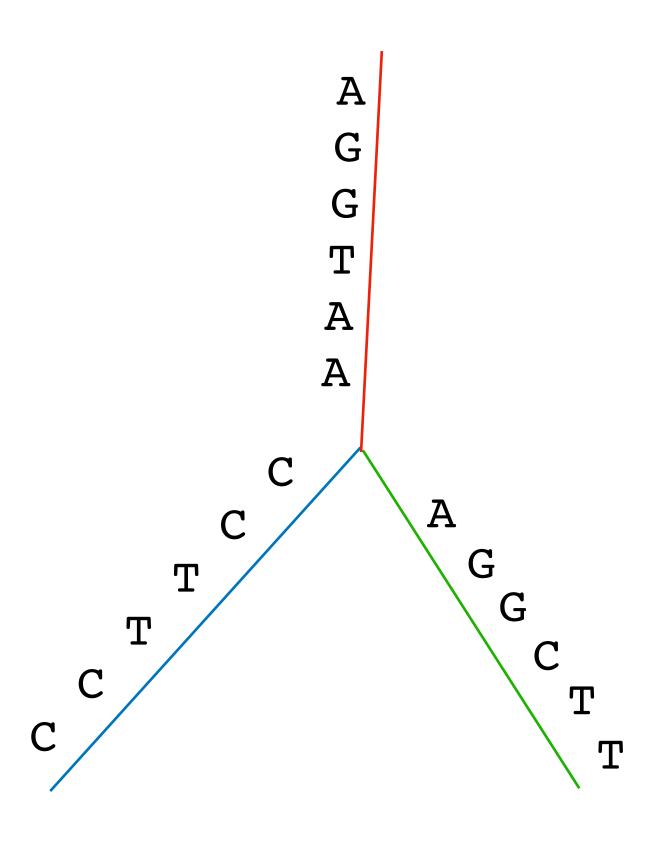
We mentioned that we want the semi-global alignment of each read to the genome, ignoring any deletions (from the genome) at the start or end of the alignment

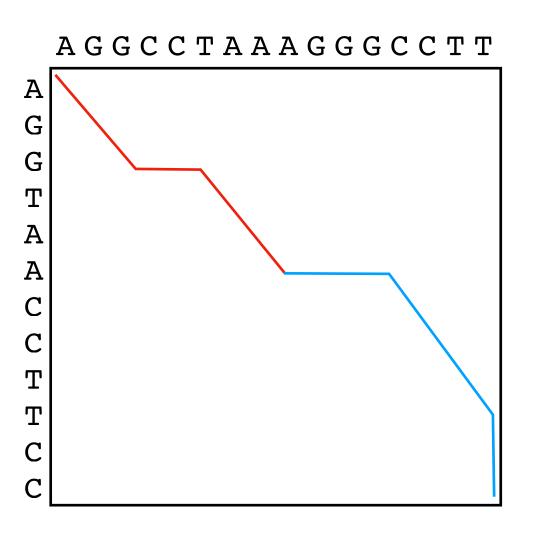
We can align the read along the suffix tree of the genome, where each row is a position in the genome

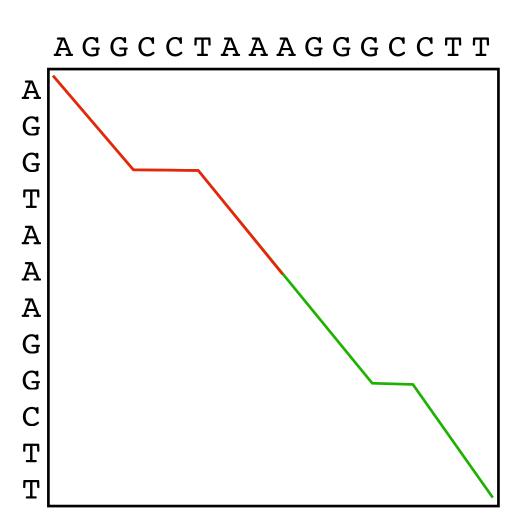
parts of the alignment matrix will be shared.

Aligning reads

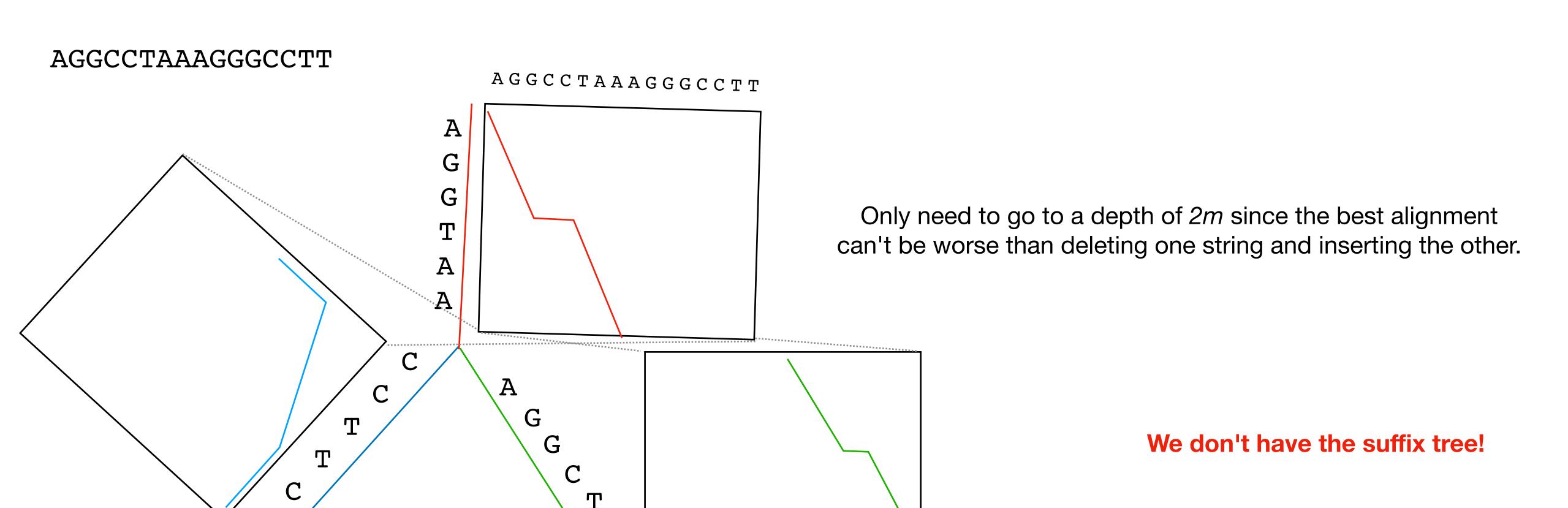
AGGCCTAAAGGGCCTT







Aligning reads



Dynamic Programming using a BWT

Since we want to save the previous rows

- we can read the characters one-by-one from the sequence
- when you reach the max depth, backtack up the tree to the last branch
- overwrite the new rows with the characters read down the new branch

How do we backtrack on a BWT?

Dynamic Programming using a BWT

```
define Branch(d,[i...j]):

for c \in idx.enumerateRight(i,j) do

process (c,d) \leftarrow

if d = 2m and score > threshold do

output alignment

if d < 2m do

Branch(d+1,idx.extendRight(c, [i,j]))

O(m\sigma)-time

O(m^2+m\sigma)-space
```

Prefix Pruning

The full dynamic program is still slow

What if we go back to hamming distance, but still use the BWT

Prefix Pruning

```
define Branch(d,k,[i...j]):
   for c ∈ idx.enumerateRight(i,j) do
       if p[d] \neq c do
          k = k-1
       if k \ge 0 do
          if d = m do
            output locations in [i,j]
         else
            Branch(d+1, k, idx.extendRight(c,[i,i]))
```

 $O(m\sigma)$ -time $O(m\sigma)$ -space

Approximate Overlaps

If we're not given a reference genome, we are left to do de novo assembly.

The first step is known as overlap mapping.

Given a set of reads $\mathbf{R} = \{R^1, R^2, ..., R^d\}$ find the set of suffix-prefix overlaps (R^i, R^j, o^{ij}) .

• Search is performed using the **reverse** BWT for $T = R^1 \$^1 R^2 \$^2 R^3 \$^3 ... R^d \$^d \$$

Paired-end reads

We mentioned previously that many times reads come in pairs from the sequencer

These pairs can be used to determine exactly where a read maps (in the case of reads that can be placed in multiple locations)

Mapping can be done independently (useful for large scale change detection), but can also be performed together

Search performed using the suffix array of the genome, which is large

Split alignment of reads

In RNA-Sequencing (RNA-Seq) the reads will have the introns removed, meaning there will be large gaps in the mapping position on the genome

If we have a complete *transcriptome* (all possible spliced transcripts), we could map reads to that, but we may not have it

There is no clean solution to this, possibilities include:

- find all error free regions of a read, piece them together if the distances are reasonable
- predict exon boundaries in the read, then align those contiguous regions

Langmead, Trapnell, Pop, Salzberg 2009

Software



Ultrafast and memory-efficient alignment of short DNA sequences to the human genome

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FM-Index

The "BWT Index" discussed previously is also called the "FM Index"

Originally defined by Ferragina and Manzini in 2000/2005

Reminder that the BWT/FM-index is:

- A data structure for a sting T containing
 - BWT_{T\$} encoded as a wavelet tree
 - an integer array C continuing the counts of each character from Σ in T

Refresher on BWTs

Can be constructed using the last character of the lexicographic order of all cyclic rotations of the text

Encodes the original text, which can be recovered by a walk in the sequence

Searching for patterns is done back to front using similar techniques to sequence recovery

Using a BWT to Align Reads

The BWT and FM-Index are insufficient for aligning reads since it doesn't allow for errors

Previously mentioned some method to overcome this

Bowtie assumes all changes are single point changes (i.e. mismatches only)

- They use a backtracking search to find matching locations
- The quality scores are used to prioritize alignments
- Other speed-ups are included to ensure all matching locations are found

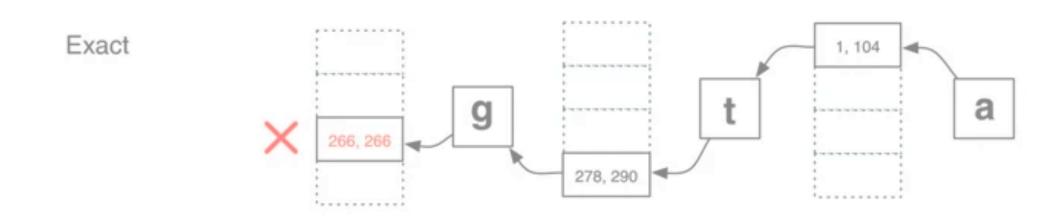
Backtracking

Start by matching the exact sequence

If the algorithm reaches a point with no matches swap out characters already matched and restart search from that there

When ties occur, start with the character with the lowest quality score, keep the rest in a stack

Keep track of how many changes are made



Backtracking Options

The user specifies the sum of the quality scores that can be changed

- this means that a mapping can have lots of low quality replacements, or
- one medium quality change

Bowtie outputs the first valid alignment by default (within the specified constraints)

- can be modified to complete the backtracking and return the "best" alignment
- •2x-3x slower to do this

User can specify a number of alignments to consider

- default is to use only one
- might want the two best alignments
- 2 alignments is ~2x slower than using only 1

Excessive Backtracking

In low quality reads, lots of time may be spent backtracking since there are many possible changes at low quality positions.

They mitigate this by creating two indexes (as we saw previously), one for the forward and one for the reverse of the string

 the backtracking is performed somewhat simultaneously on both index as we will see next

One other step they take is to concentrate on the "high-quality" end of a read (the first 28 characters read) which is most reliable

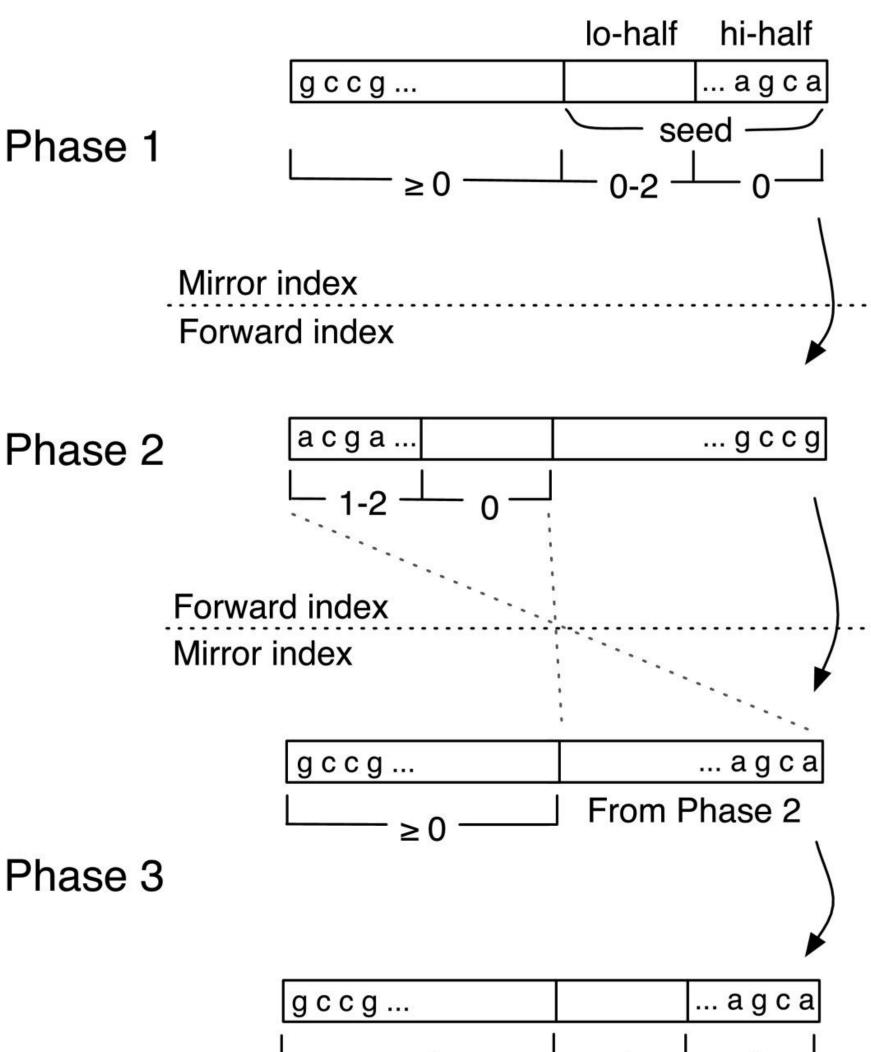
Phased Search

Split the seed (first 28 bases) into two parts, hi-half and lohalf

Assume we're allowing 2 changes in the seed, a good alignment will have either:

1. no mismatches acga.. Phase 2 2. no mismatches in hi-half, 1 or 2 mismatches in lo-half 3. 1 or 2 mismatches in hi-half, no mismatches in lo-half Forward index 4. 1 mismatch in hi-half, 1 mismatch in lo-half Mirror index gccg... Phase 1 uses the mirror index and invokes the aligner to find alignments for cases 1 & 2. Phase 2 finds partial alignments with mismatches only in the hi-half, and gccg... phase 3 attempts to extend those partial alignments into full alignments. Finally, phase 3 invokes the aligner to find alignments for case 4.

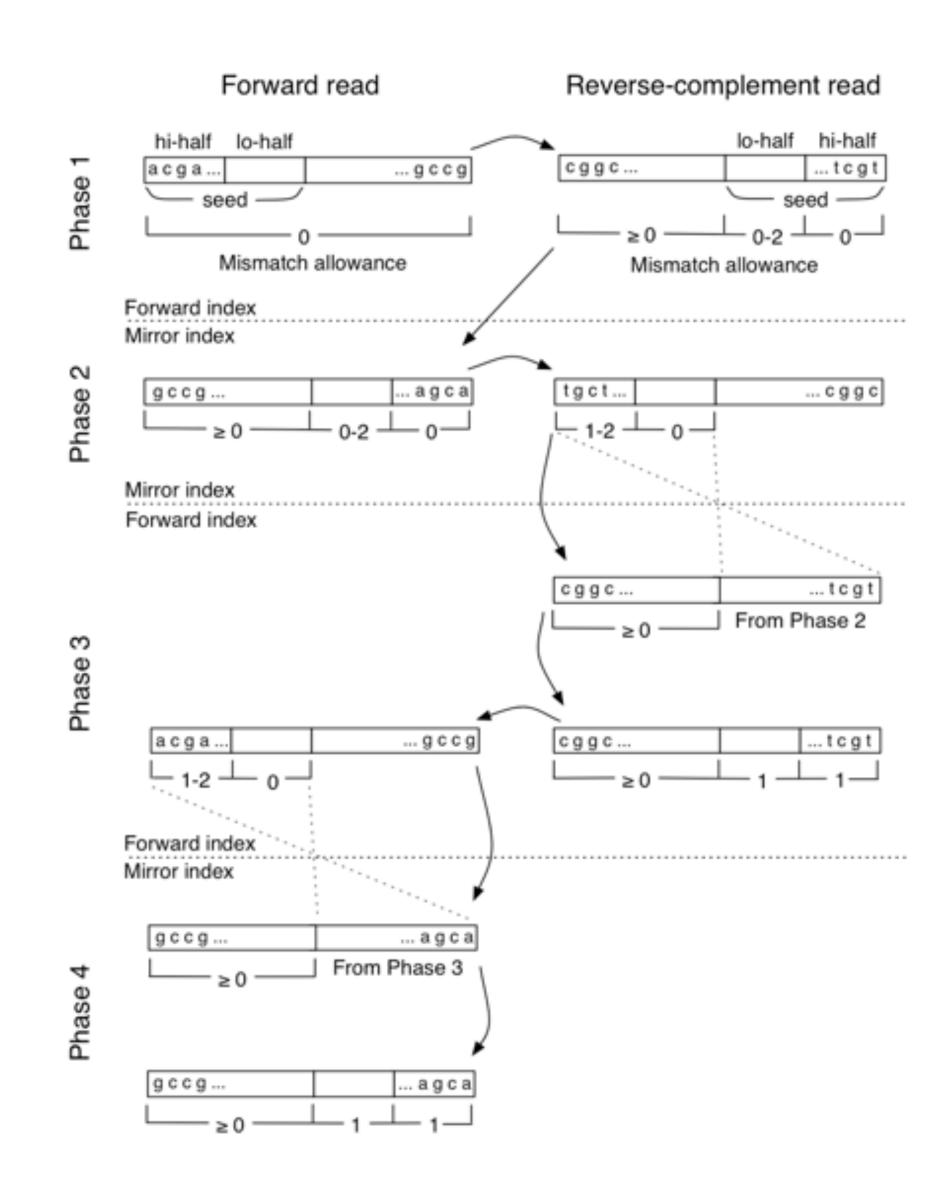
Phases 2 and 3 cooperate to find alignments for case 3:



Phased search with reverse strand

Since both the read and its reverse complement are possibilities for ma match, need to consider both.

Phases 2-4 here map to phases 1-3 previously.



Performance

Maq (Li, Ruan, Durban 2008), SOAP (Li, Li, Kristiansen, Wang 2008) the leading competitors at the time

Both used hashing to find potential mapping locations

Performance

	Platform	CPU time	Wall clock time	Reads mapped per hour (millions)	Peak virtual memory footprint (megabytes)	Bowtie speed-up	Reads aligned (%)
Bowtie -v 2	Server	15 m 7 s	15 m 41 s	33.8	1,149	351×	67.4
SOAP		91 h 57 m 35 s	91 h 47 m 46 s	0.10	13,619		67.3
Bowtie	PC	16 m 41 s	17 m 57 s	29.5	1,353	59.8×	71.9
MAQ		17 h 46 m 35 s	17 h 53 m 7 s	0.49	804		74.7
Bowtie	Server	17 m 58 s	18 m 26 s	28.8	1,353	107×	71.9
MAQ		32 h 56 m 53 s	32 h 58 m 39 s	0.27	804		74.7

Take Aways

Bowtie was (at the time) the fastest short read aligniner

Used a one-time index based on a BWT that could be reused (novel at the time)

Is able to run on a standard PC

When first published didn't use mate-pair information



```
Read (reverse complement)
Read
CCAGTAGCTCTCAGCCTTATTTTACCCAGGCCTGTA TACAGGCCTGGGTAAAATAAGGCTGAGAGCTACTGG
                      Policy: extract 16 nt seed every 10 nt
Sccds
+, 0: CCAGTAGCTCTCAGCC
                                        -, 0: TACAGGCCTGGGTAAA
                                                 -, 10: GGTAAAATAAGGCTGA
         +, 10: TCAGCCTTATTTTACC
                                                          -, 20: GGCTGAGAGCTACTGG
                   +, 20: TTTACCCAGGCCTGTA
                                                Seed alignments
Seeds
                               Ungapped
                                                (as Burrows-Wheeler ranges)
 +, 0: CCAGTAGCTCTCAGCC
                              alignment with
                                                 [211, 212], [212, 214] }
                                FM Index
+, 10: TCAGCCTTATTTTACC
                                                 [653, 654], [651, 653] }
+, 20: TTTACCCAGGCCTGTA
                                                 [684, 635] }
 -, 0: TACAGGCCTGGGTAAA
                                 acces.
                                 accinca.
-, 10: GGTAAAATAAGGCTGA
-, 20: GGCTGAGAGCTACTGG
                                                  [624, 625] )
 Seed alignments (as BW ranges)
                                                     Extension candidates
                                     Walk-left with
 { [211, 212], [212, 214] }
                                       FM Index
                                                     BW row: 684: chr12:1955
 { [653, 654], [651, 653] }
                                                     BW row: 624: chr2:462
 { [684, 685] }
                                                  → BW row:211: chr4:762
                                        ACTUAL S
                       Prioritize
                                                     BW row:213: chr12:1935
                                        acqlicta
                                                     BW row:652: chr12:1945
 { [624, 625] }
                                        g > 4 = 1 = c
 Extension cancidates
                                          SAM alignments
                            SIMD dynamic
                            programming
                                           rl 0 chr12
                                                           1936
BW row: 684: chr12:1955
                               aligner
BW row: 624: chr2:462
                                               CCAGTAGCTCTCAGCCTTATTTTACCCAGGCCTGTA
                                              BW row:211: chr4:762
                                                      XS:i:-2 XN:i:0
                                               AS:1:0
BW row:213: chr12:1935
                                                       X0:i:0 XG:i:0
                                               XM:1:0
                                                       MD:Z:36 YT:Z:UU
                                               NM:i:0
BW row: 652: chr12:1945
                                               YM:1:0
```

